



Fast Analysis of $\Delta 8$ -THC, $\Delta 9$ -THC, and Isomeric Hydroxy and Carboxy Metabolites in Whole Blood by LC-MS/MS

By Haley Berkland

Introduction

The testing of whole blood samples for tetrahydrocannabinol ($\Delta 9$ -THC) consumption is routine and has been performed for many decades. Since $\Delta 9$ -THC is metabolized into 11-hydroxy- $\Delta 9$ -tetrahydrocannabinol (11-OH- $\Delta 9$ -THC) and further into 11-nor-9-carboxy- $\Delta 9$ -THC ($\Delta 9$ -THC-COOH), it is important to test for the parent compound and both metabolites to properly monitor for THC usage.

As more isomers of $\Delta 9$ -THC become available on the market, testing has become more complicated, and novel methods are needed to achieve isomeric resolution. One such isomer, $\Delta 8$ -THC, is federally unregulated in the United States and readily available for purchase in many stores. This compound forms its own hydroxylated and carboxylated metabolites, (11-OH- $\Delta 8$ -THC and $\Delta 8$ -THC-COOH), that must be resolved from their isomeric $\Delta 9$ counterparts. Chromatographic resolution of all three isomeric pairs is essential for reporting accurate clinical specimen results, and poor resolution, especially when one isomer is present in much greater abundance than the other, can cause invalid data.

To improve reporting accuracy, the following method was developed to adequately resolve all six isomers in whole blood. Linearity, accuracy, precision, and the potential for cross-analyte interferences were assessed.

Related Products

- *Raptor FluoroPhenyl column (cat.# 9319A1E)*
- *Raptor FluoroPhenyl EXP guard column cartridge (cat.# 9319A0253)*
- *$\Delta 8$ -Tetrahydrocannabinol standard (cat.# 34090)*
- *$\Delta 9$ -Tetrahydrocannabinol standard (cat.# 34067)*
- *(\pm)11-nor-9-carboxy- $\Delta 9$ -THC standard (cat.# 34068)*
- *2 mL vial (cat.# 21143)*
- *Vial insert (cat.# 21776)*
- *Short-cap, screw-vial closure (cat.# 24498)*

Experimental

Liquid-Liquid Extraction [1]

1. 500 μ L aliquots of blank whole blood were added to glass test tubes.
2. 50 μ L of calibrator or QC standard was added to each tube. 50 μ L of internal standard (1000 ng/mL of $\Delta 9$ -THC-D3; 11-OH- $\Delta 9$ -THC-D3; and $\Delta 9$ -THC-COOH-D3) was added to each tube and vortexed.
3. 500 μ L of HPLC grade water was added to each tube and vortexed.
4. 100 μ L of 1N HCl was added to each tube and vortexed.
5. 2.5 mL of 80:20 hexanes:ethyl acetate was added to each tube, capped, and vortexed until visibly combined.
6. Samples were centrifuged at 4200 rpm for 15 minutes or until the two layers had completely separated.
7. The supernatant was pipetted off each sample, transferred to a clean test tube, and dried down under nitrogen.
8. Samples were reconstituted in 100 μ L of 50:50 water:methanol (both containing 0.1% formic acid), vortexed, and transferred to an LC vial with an insert.

Instrument Parameters

The LC-MS/MS method detailed in Tables I and II was developed on a Raptor FluoroPhenyl column in order to optimally separate Δ^8 -THC; Δ^9 -THC; 11-OH- Δ^8 -THC; 11-OH- Δ^9 -THC; Δ^8 -THC-COOH; and Δ^9 -THC-COOH. MRM transitions and ESI mode for each analyte are given in Figure 1.

Table I: Chromatography Gradient

Time (min)	Flow Rate (mL/min)	%A	%B
0.00	0.8	36	64
6.50	0.8	36	64
6.60	0.8	32	68
13.00	0.8	32	68
13.10	0.8	0	100
14.00	0.8	0	100
14.10	0.8	36	64
16.00	0.8	36	64

Table II: LC Method Parameters

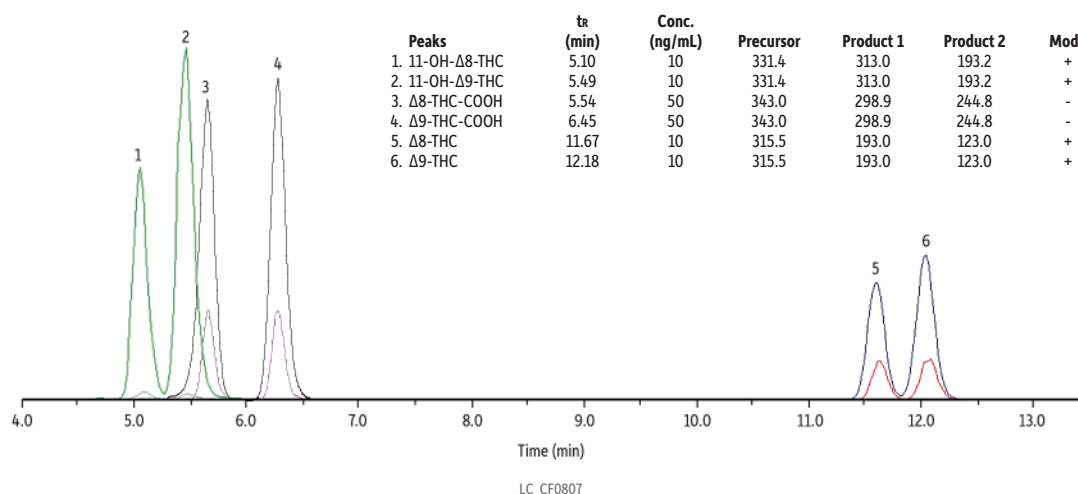
Column	Raptor FluoroPhenyl, 2.7 μ m, 100 mm x 3.0 mm ID (cat.# 9319A1E)
Guard	Raptor FluoroPhenyl EXP guard column cartridge 5 mm, 3.0 mm ID, 2.7 μ m (cat.# 9319A0253)
Column Temperature	40 °C
Mobile Phase A	0.1% formic acid in water
Mobile Phase B	0.1% formic acid in methanol
Injection Volume	10 μ L

Results & Discussion

Chromatographic Performance

As shown in Figure 1, all three sets of isomers (six total analytes) were well separated in a fast 13-minute gradient (16-minute total analysis time) on a Raptor FluoroPhenyl column. The FluoroPhenyl stationary phase provided better selectivity for all three isomer pairs compared to alternative column chemistries, such as biphenyl or C18.

Figure 1: Separation of $\Delta 8/9$ -THC, Hydroxy, and Carboxy Metabolites in Whole Blood



Column: Raptor FluoroPhenyl (cat.# 9319A1E); Dimensions: 100 mm x 3.0 mm ID; Particle Size: 2.7 μ m; Pore Size: 90 Å; Guard Column: Raptor FluoroPhenyl EXP guard column cartridge 5 mm, 3.0 mm ID, 2.7 μ m (cat.# 9319A0253); Temp.: 40 °C; **Standard/Sample:** $\Delta 8$ -Tetrahydrocannabinol ($\Delta 8$ -THC) (cat.# 34090); $\Delta 9$ -Tetrahydrocannabinol ($\Delta 9$ -THC) (cat.# 34067); (\pm)11-nor-9-carboxy- Δ -9-THC ($\Delta 9$ -THC-COOH) (cat.# 34068); Other compounds obtained separately.; Diluent: 50:50 Methanol:water, both with 0.1% formic acid; Inj. Vol.: 10 μ L; **Mobile Phase:** A. Water, 0.1% formic acid; B. Methanol, 0.1% formic acid. **Gradient (%B):** 0.00 min (64%); 6.50 min (64%); 6.60 min (68%); 13.00 min (68%); 13.10 min (100%); 14.00 min (100%); 14.10 min (64%); 16.00 min (64%). **Flow:** 0.8 mL/min; Max Pressure: 440 bar; **Detector:** SCIEX 4500 MS/MS; Ion Source: Electrospray; Ion Mode: ESI+/ESI-; **Sample Preparation** 500 μ L of whole blood was transferred to a 15 mL glass test tube, 50 μ L of internal standard and 50 μ L of control material were transferred to the test tube and vortexed. 500 μ L of HPLC grade water was added to each sample and vortexed. 100 μ L of 1N HCl was added to each sample and vortexed. 2.5 mL of 80:20 hexanes:ethyl acetate was added to each sample and vortexed until visibly combined. Samples were centrifuged at 4200 rpm for 15 minutes. The top layer was transferred to a new glass test tube and dried down under nitrogen. Samples were reconstituted with 100 μ L of 50:50 methanol:water, both containing 0.1% formic acid, and vortexed. Samples were transferred to 2 mL screw-thread vials (cat.# 21143) with glass inserts (cat.# 21776) and capped with short-cap, screw-vial closures (cat.# 24498).

Linearity, Accuracy, and Precision

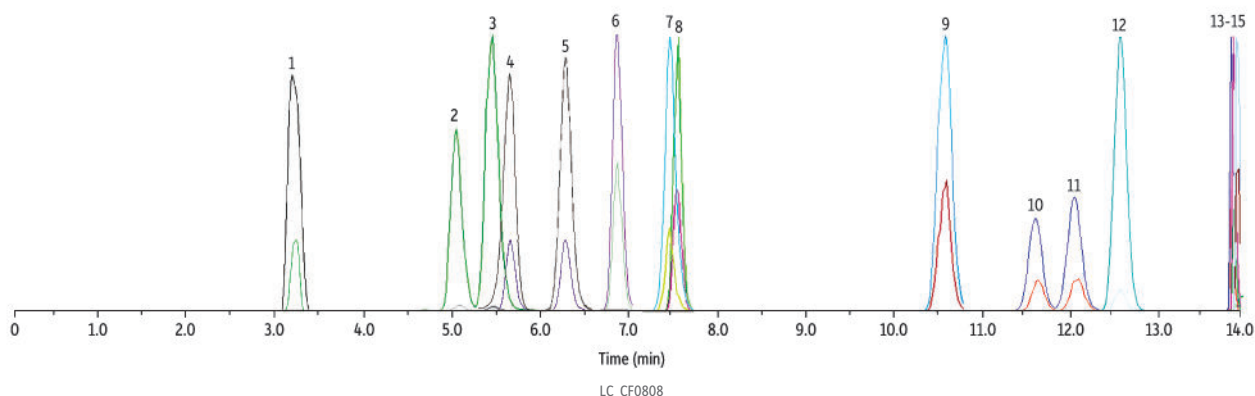
Linearity was demonstrated using a $1/x^2$ weighted linear regression, and all analytes showed acceptable R^2 values (≥ 0.99). The linear calibration ranges were 0.5–100 ng/mL for 11-OH- $\Delta 8$ -THC; 11-OH- $\Delta 9$ -THC; $\Delta 8$ -THC; and $\Delta 9$ -THC and 2.5–500 ng/mL for $\Delta 8$ -THC-COOH and $\Delta 9$ -THC-COOH.

Accuracy and precision were assessed on three different days using low, mid, and high QC samples spiked at 5, 10, and 50 ng/mL for 11-OH- $\Delta 8$ -THC; 11-OH- $\Delta 9$ -THC; $\Delta 8$ -THC; and $\Delta 9$ -THC and at 25, 50, and 250 ng/mL for $\Delta 8$ -THC-COOH and $\Delta 9$ -THC-COOH. Method accuracy was demonstrated by recovery values being within 10% of the nominal concentrations for the QC samples at all levels. The %RSD was under 20% for both intraday and interday testing, indicating acceptable method precision.

Cross-Analyte Interferences

To demonstrate that other naturally occurring cannabinoids would not interfere with the target compounds, we spiked whole blood with nine commonly encountered and/or structurally similar cannabinoids: CBDV, CBD, CBG, THCV, exo-THC, CBL, CBN, $\Delta 10$ -THC, and CBC. All nine cannabinoids were fully resolved from the analytes of interest on the Raptor FluoroPhenyl column and no cross-analyte interferences are expected (Figure 2).

Figure 2: Separation of $\Delta 8/9$ -THC, Hydroxy, and Carboxy Metabolites from Nine Cannabinoids



Peaks	tr (min)	Conc. (ng/mL)	Precursor	Product 1	Product 2	Mode
1. CBDV	3.52	50	287.4	165.0	231.1	+
2. 11-OH- $\Delta 8$ -THC	5.09	50	331.4	313.0	193.2	+
3. 11-OH- $\Delta 9$ -THC	5.50	50	331.4	313.0	193.2	+
4. $\Delta 8$ -THC-COOH	5.53	250	343.0	298.9	244.8	-
5. $\Delta 9$ -THC-COOH	6.44	250	343.0	298.9	244.8	-
6. CBD	6.81	50	315.5	193.0	259.0	+
7. CBG	7.56	50	317.5	193.0	123.0	+
8. THCV	7.65	50	287.4	165.0	123.0	+
9. exo-THC	10.63	50	315.5	193.0	259.0	+
10. $\Delta 8$ -THC	11.67	50	315.5	193.0	123.0	+
11. $\Delta 9$ -THC	12.17	50	315.5	193.0	123.0	+
12. CBL	12.43	50	315.5	235.2	193.0	+
13. $\Delta 10$ -THC	13.88	50	315.5	193.1	259.0	+
14. CBN	13.93	50	311.4	223.0	293.1	+
15. CBC	13.93	50	315.5	193.1	259.0	+

Column: Raptor FluoroPhenyl (cat.# 9319A1E); Dimensions: 100 mm x 3.0 mm ID; Particle Size: 2.7 μ m; Pore Size: 90 Å; Guard Column: Raptor FluoroPhenyl EXP guard column cartridge 5 mm, 3.0 mm ID, 2.7 μ m (cat.# 9319A0253); Temp.: 40 °C; **Standard/Sample:** Cannabinoids Neutrals 9 standard (cat.# 34132); $\Delta 8$ -Tetrahydrocannabinol ($\Delta 8$ -THC) (cat.# 34090); $\Delta 9$ -Tetrahydrocannabinol ($\Delta 9$ -THC) (cat.# 34067); (\pm)11-nor-9-carboxy- Δ -9-THC ($\Delta 9$ -THC-COOH) (cat.# 34068); Other compounds obtained separately; Diluent: 50:50 Methanol:water, both with 0.1% formic acid; Inj. Vol.: 10 μ L; **Mobile Phase:** A. Water, 0.1% formic acid; B. Methanol, 0.1% formic acid. **Gradient (%B):** 0.00 min (64%); 6.50 min (64%); 6.60 min (68%); 13.00 min (68%); 13.10 min (100%); 14.00 min (100%); 14.10 min (64%); 16.00 min (64%). **Flow:** 0.8 mL/min; Max Pressure: 440 bar; **Detector:** SCIEX 4500; Ion Source: Electrospray; Ion Mode: ESI+/ESI-; **Sample Preparation** 500 μ L of whole blood was transferred to a 15 mL glass test tube. 50 μ L of internal standard and 50 μ L of control material were transferred to the test tube and vortexed. 500 μ L of HPLC grade water was added to each sample and vortexed. 100 μ L of 1N HCl was added to each sample and vortexed. 2.5 mL of 80:20 hexanes:ethyl acetate was added to each sample and vortexed until visibly combined. Samples were centrifuged at 4200 rpm for 15 minutes. The top layer was transferred to a new glass test tube and dried down under nitrogen. Samples were reconstituted with 100 μ L of 50:50 methanol:water, both containing 0.1% formic acid, and vortexed. Samples were transferred to 2 mL screw-thread vials (cat.# 21143) with glass inserts (cat.# 21776) and capped with short-cap, screw-vial closures (cat.# 24498).

Conclusion

The fast LC-MS/MS method established here on a Raptor FluoroPhenyl column provides essential chromatographic resolution of $\Delta 8$ -THC and $\Delta 9$ -THC isomers as well as their isomeric hydroxy and carboxy metabolites in whole blood. In addition, the target compounds were completely separated from potentially interfering cannabinoids, helping ensure that labs can report more accurate clinical results.

References

1. N.B. Tiscione, R. Miller, X. Shan, J. Sprague, D.T. Yeatman, An efficient, robust method for the determination of cannabinoids in whole blood by LC-MS-MS, J. Ana. Toxicol. 40(8) (2016) 639-648. <https://doi.org/10.1093/jat/bkw063>

Raptor FluoroPhenyl HPLC Columns

- Retains hydrophobic, polar, and aromatic compounds.
- Has orthogonal selectivity to a C18.
- Exceptionally reproducible—predictable performance from every column.
- Part of Restek's Raptor LC column line featuring 1.8, 2.7, and 5 µm SPP core-shell silica.
- Switch to a Raptor FluoroPhenyl LC column when you need more retention and selectivity for basic and hydrophilic compounds than you can achieve on a C18.



Catalog No.	Particle Size	Internal Diameter (ID)	Length	Units
9319A1E	2.7 µm	3.0 mm	100 mm	ea.

Raptor FluoroPhenyl EXP Guard Column Cartridge

- Free-Turn architecture lets you change cartridges by hand without breaking inlet/outlet fluid connections—no tools needed.
- Patented titanium hybrid ferrules can be installed repeatedly without compromising high-pressure seal.
- Auto-adjusting design provides ZDV (zero dead volume) connection to any 10-32 female port.
- Guard column cartridges require EXP direct connect holder (cat.# 25808).
- Pair with EXP hand-tight fitting (cat.# 25937–25938) for tool-free installation.



Catalog No.	Particle Size	Internal Diameter (ID)	Length	Units
16094	2.7 µm	3.0 mm	5 mm	3-pk.

Delta 8-Tetrahydrocannabinol (Delta 8-THC) Standard

Catalog No.	Concentration	Solvent	Volume	Unit
34090	1000 µg/mL	P&T Methanol	1 mL/ampul	ea.

Delta 9-Tetrahydrocannabinol (Delta 9-THC) Standard

Catalog No.	Concentration	Solvent	Volume	Unit
34067	1000 µg/mL	Methanol	1 mL/ampul	ea.



(±)11-nor-9-carboxy-Delta-9-THC Standard

Catalog No.	Concentration	Solvent	Volume	Unit
34068	100 µg/mL	Methanol	1 mL/ampul	ea.



2.0 mL Vial

Catalog No.	Product Name	Units
21143	Short-Cap Vial with Grad Marking Spot, 9-425 Screw-Thread, 2.0 mL, 9 mm, 12 x 32 (vial only), Amber	1000-pk



Vial Insert

Catalog No.	Product Name	Units
21776	Vial Inserts, Glass, Big Mouth w/Bottom Spring, 250 µL	100-pk.



Short-cap, screw-vial closure

Catalog No.	Product Name	Units
24498	Short Screw Cap, Polypropylene, Screw-Thread, PTFE/Silicone/PTFE Septa, Blue, Preassembled, 2.0 mL, 9 mm	1000-pk.