

S. K. Ruiz Perez^a, E. Ceccon^b, F. Rodriguez Carrero^c, D. Bell^d, P. Conolly^d;

Introduction

Multi-methods enable analysis of more components in a single run, saving instrument time and reducing solvent waste and energy consumption. However, the growing range of analytes, especially **polar compounds**, increases demands on the stationary phase. The traditional C18 phase is reaching its limits, making versatile alternatives essential.

Free virtual tools like the **Pro EZLC Chromatogram Modeler** and *Method Translator* further reduce lab time, energy use, and instrument wear by enabling method development without lab work.

The **versatile Biphenyl** phase can undergo different separation mechanisms (Figure 1):

-
- The figure compares the chemical structures and photophysical properties of two dyes: Biphemyl and C18. On the left, the Biphemyl dye is shown with its chemical structure, which includes a biphenyl core and a trimethylsilyl group. A label 'free rotation' points to the biphenyl bond. Below the structure is a circular diagram divided into four quadrants: Disposition (blue), H-Bonding (green), Polarizability (red), and Cation Exchange (orange). On the right, the C18 dye is shown with its chemical structure, which includes a C18 core and a trimethylsilyl group. Below the structure is a similar circular diagram with the same four quadrants. A vertical dashed line separates the two diagrams, with 'VS.' written above it. The C18 dye's diagram shows a different distribution of properties compared to the Biphemyl dye.

This duality in separation mechanism and the 100% water compatibility makes it particularly useful for **isomeric isobars, polar compounds, MS-detection.**

Acetonitrile	Methanol
Mobile phase: 0.5% formic acid in water (pH 2.25); ACN 0.1% formic acid (1:1) Flow rate: 1.0 mL/min, isocratic UV detection: @254 nm Temperature: room temperature Injection volume: 5 µL with 100 µg/mL	Mobile phase: 0.5% formic acid in water (pH 2.25); MeOH 0.1% formic acid Flow rate: 1.0 mL/min, isocratic UV detection: @254 nm Temperature: room temperature Injection volume: 5 µL with 100 µg/mL
Classic C18 5 µm 4.6 mm x 150 mm	Classic C18 5 µm 4.6 mm x 150 mm
Biphenyl 5 µm 4.6 mm x 150 mm	Biphenyl 5 µm 4.6 mm x 150 mm
Similar Selectivity	Orthogonal Selectivity

Examples for Multi-Methods

[illegible]

Mycotoxins

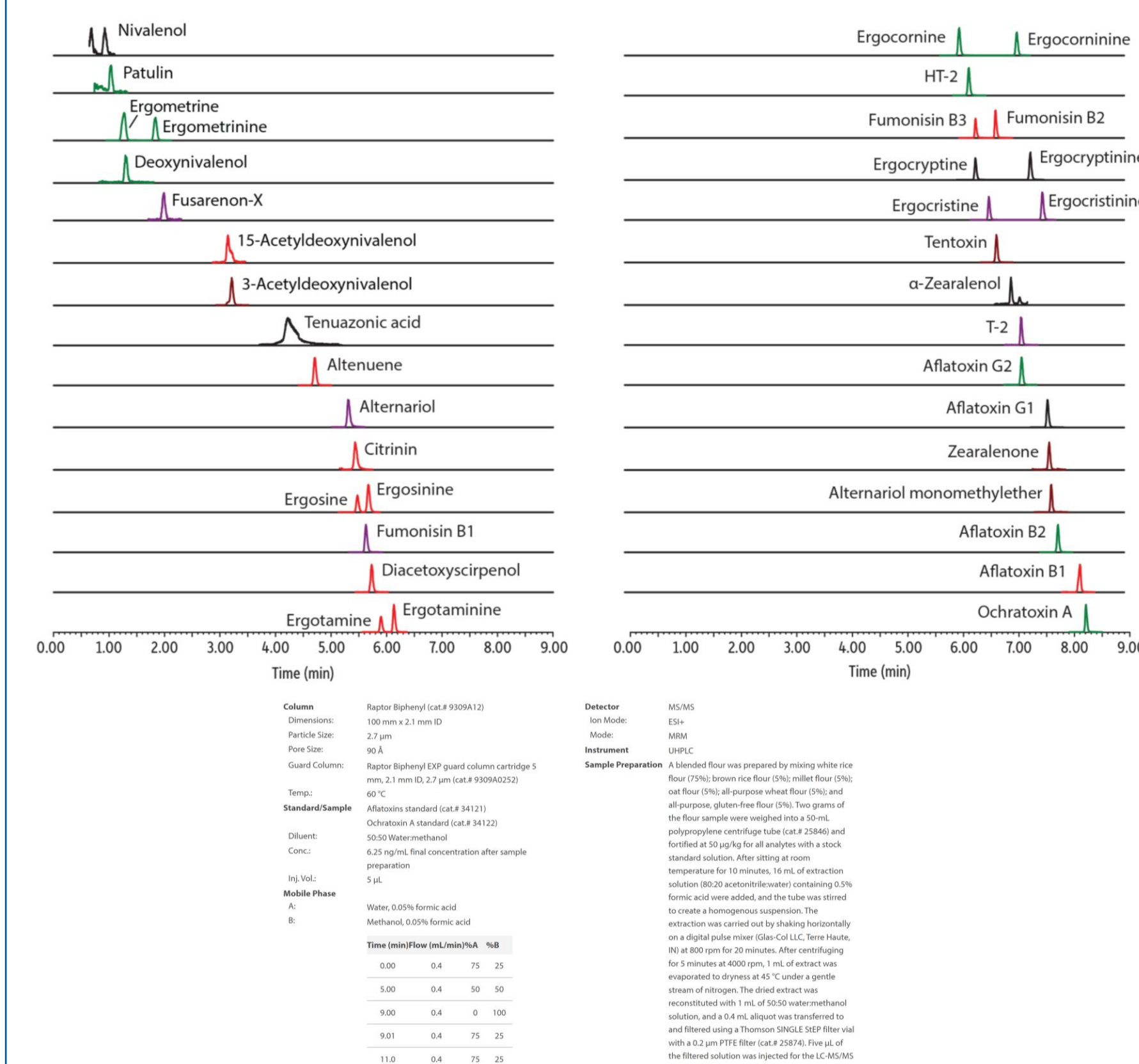
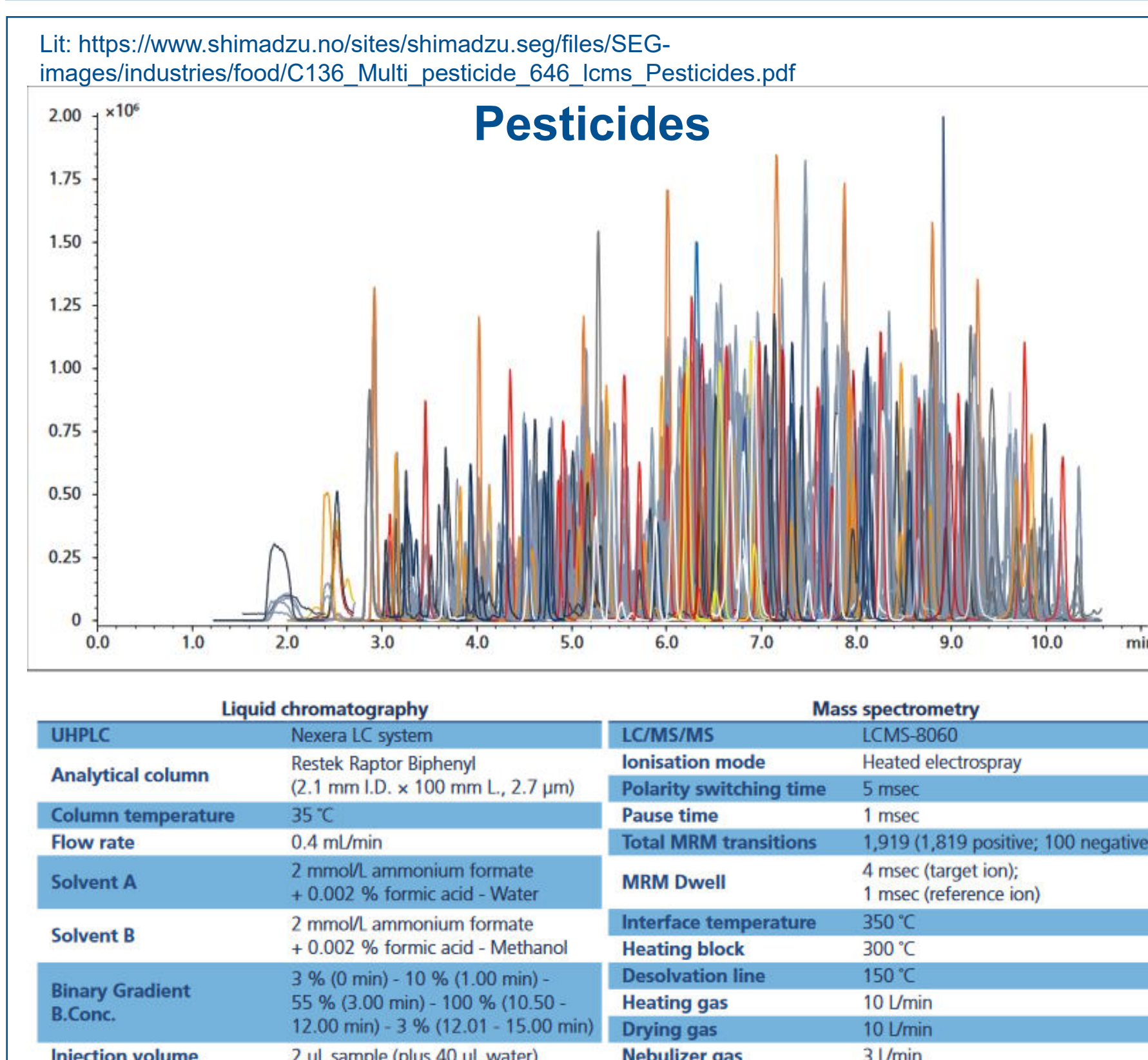


Figure 4 – Chromatogram and method conditions for our **Ergot alkaloids, Alternaria toxins and other major Mycotoxins** in fortified flour sample on Raptor Biphenyl by LC-MS/MS.



Virtual Method Development

Pro EZLC Chromatogram

Compounds

Conditions

My EZLC

Compounds

Conditions

My EZLC

Compound Class:

Drugs of Abuse

Compound Name

Search

1-4-Ephedrine

221-98-2

✓

α-Hydroxyphenylacetamide

37115-43-8

✓

α-Hydroxymisclizolam

59468-90-5

✓

α-Hydroxytryptolam

37115-45-0

✓

α-Pyrrolidinococaineamphetamine

13415-59-3

✓

α-Pyrrolidinococainehexanophenone

21-769-9

✓

1-(3-Chlorophenyl)pyiperazine (mCPP)

6640-24-0

✓

1-CB

76835-14-6

✓

1-Hydroxymisclizolam

56354-06-4

✓

11-Nor-carboxy-Δ⁹-THC

20971-15-3

✓

Hydroxyethylflurazepam

98068-59-4

✓

2-Methyl-AP-237

✓

291 Compounds Selected

71 Isomers Total

Select All

Clear

Target All

Clear

Phase:

Raptor Elypset

Detector:

MS

Generate Model

Length

50.00 mm

Inner Diameter

3.00 mm

Particle Size

2.70 μm

Available Columns

50, 3.00, 2.70, 1.70

Volume Effects

Dwell Volume

0.25 mL

Extra-Column Volume

25.00 μL

Mobile Phase

Eluent A

Water

0.1% Formic Acid

Eluent B

Methanol

0.1% Formic Acid

Temperature

30.00 °C

Back Pressure

psi

2862 psig

Gradient Program

☐ Add Start Isocratic Hold

Time (min)

0.00

0.00

☐ Or Gradient Steps

Time (min)

4.00

0.00

☒ Add Final Isocratic Hold

Time (min)

5.00

0.00

☐ Add Re-equilibration Time

Time (min)

5.00

0.00

Target Resolution

1.50

Minimum Gradient Steps

1

Results

Gradient Time × Delay / Run Time

5.62 / 5.62 min

To

0.21 min

Isocratic Compounds Separated

127

Critical Pair

128,129

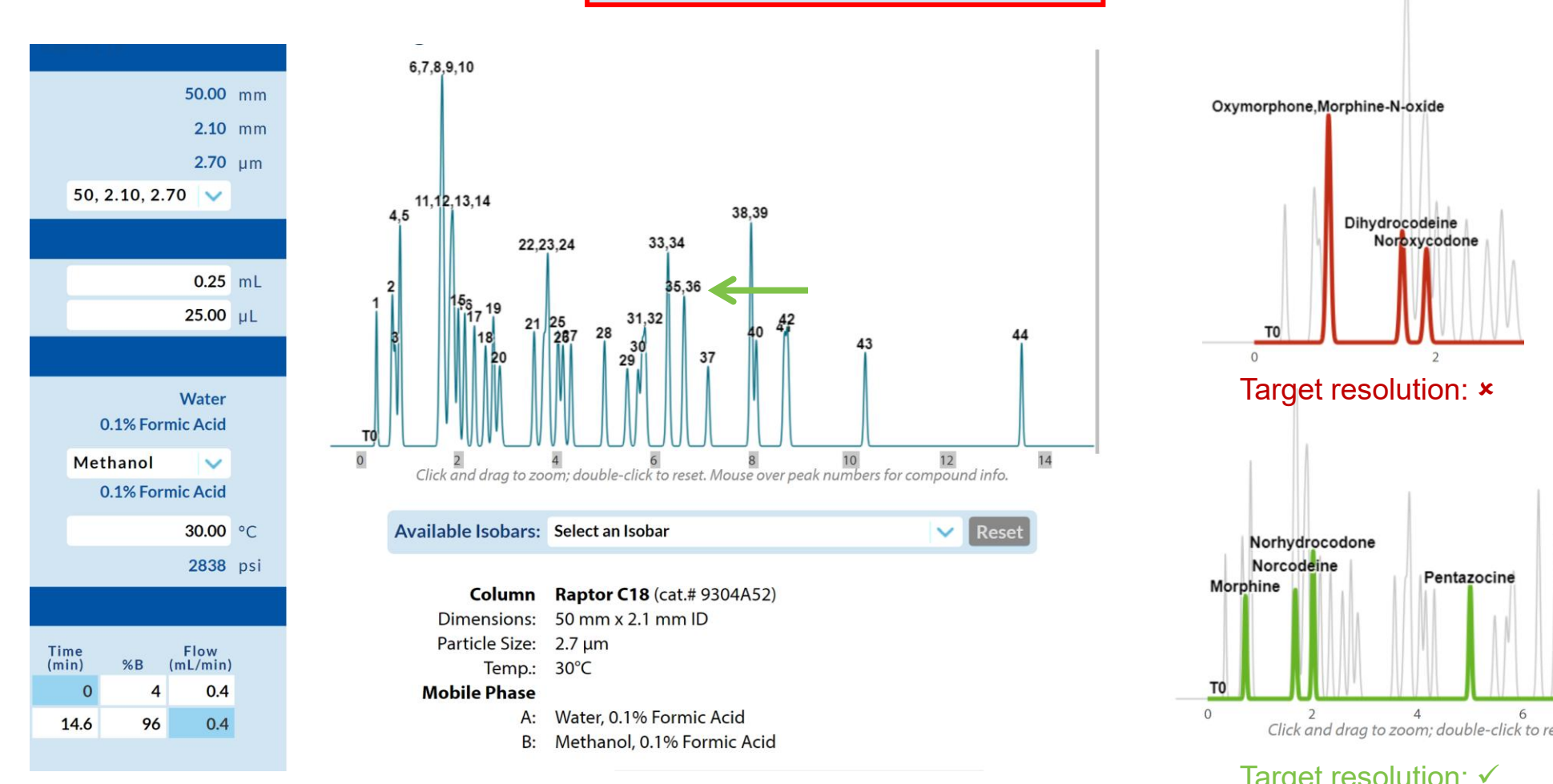
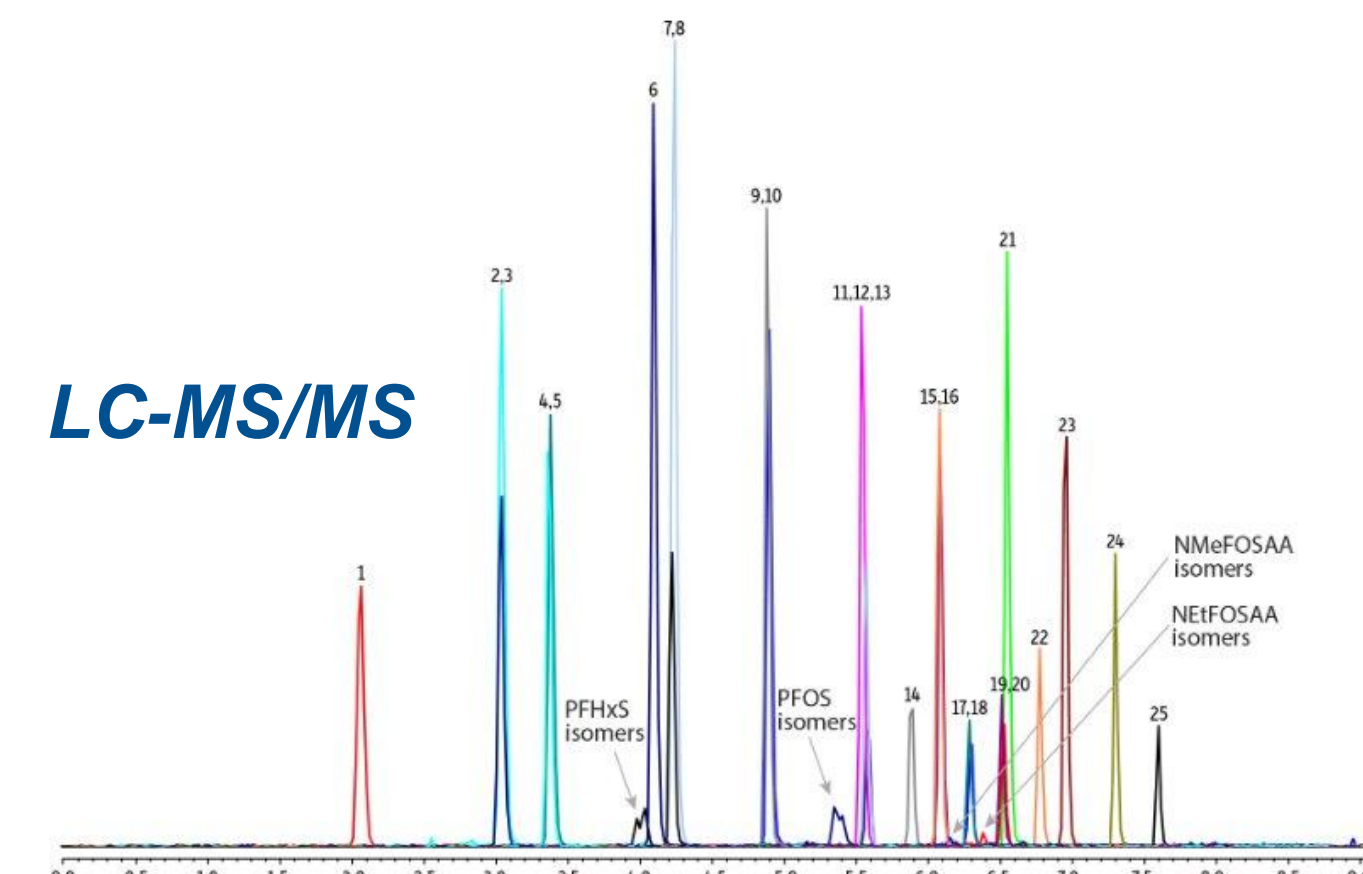
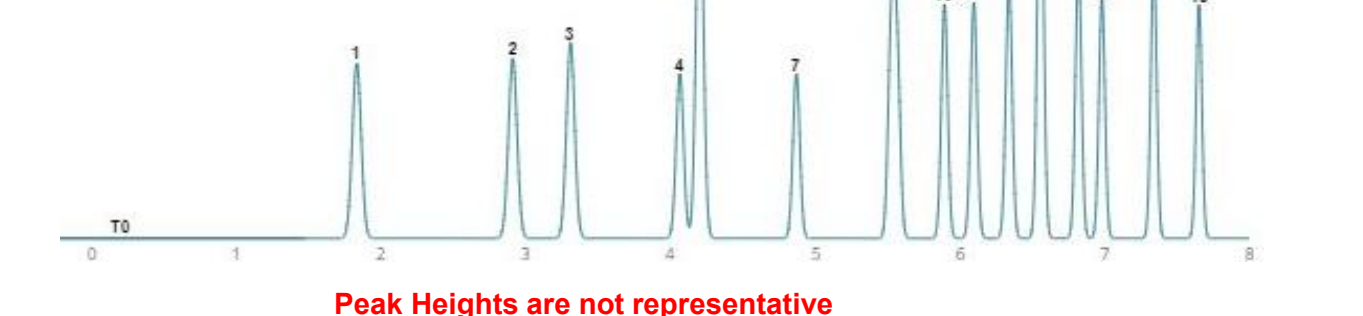


Figure 6 – From left to right: 1. Search your compounds in the libraries and see the isobars of your method, 2. adapt the conditions to your equipment and methods, 3. save your method development for futures alterations, 4. See if your critical isobars are separated or not.

Model vs. Reality

EPA 537.1

EZLC



Compound	LC_EV050 (min)	Modelled Result	
Name	RT (min)	RT (min)	Diff (min)
PFBS	2.06	1.84	-0.22
PFHxA	3.04	2.91	-0.13
HPFO-D4	3.38	3.31	-0.07
PFHpA	4.29	4.07	-0.02
PFNA	4.22	4.20	-0.02
ADONA	4.24	4.21	-0.03
PFOA	4.90	4.87	-0.03
PFNA	5.54	5.53	-0.01
PFOS	5.58	5.57	-0.01
9Cl-PF3ONS	5.88	5.89	0.01
PFDA	6.08	6.10	0.02
NMeFOSAA	6.36	6.34	0.04
NEtFOSAA	6.52	6.56	0.04
PFUnA	6.55	6.56	0.01
11Cl-PF3Ods	6.77	6.82	0.05
PFDoA	6.95	6.98	0.03
PFTfDA	7.30	7.34	0.04
PFTeDA	7.60	7.65	0.05
		Average Difference (min):	-0.01

Conclusion

Greener workflows can be implemented in any lab with minimal effort and cost. Free method development tools allow you to optimize methods virtually, saving instrument time, energy, and solvent waste. Analyzing more components in less time with a single method (multi-method) increases both efficiency and sustainability. Choosing the right stationary phase is key—columns with multiple retention mechanisms, like the Biphenyl phase, offer greater versatility and greener performance.